

# Dark fermentative hydrogen production rate from glucose using facultative anaerobe bacteria *E coli*

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**Abstract**—Hydrogen is a best and environmental- friendly energy source, its combustion results only in water as product. Hydrogen may be produced by a number of diverse processes including biological processes using dark fermentation, direct or indirect bio photolysis and photo fermentation. The higher hydrogen production rates were acquired using dark fermentation process on the contrary to other bioprocesses. Dark fermentation can be accomplished by facultative and obligate anaerobic microorganisms. In this study, the hydrogen production was investigated using facultative anaerobe bacteria *E coli*. The optimization of some key factors for biohydrogen production considerably elevates the biohydrogen production effectiveness.

**Keywords**—*Biohydrogen; Dark fermentation; E coli; Hydrogen production rate.*

## I. INTRODUCTION

Hydrogen gas is a clean and high energy fuel which can be produced in fuel cells for electricity generation. Hydrogen is considered as the major energy carrier of the future. Unlike fossil fuels, hydrogen is not available in nature and requires expensive production methods [1]. Biological production of hydrogen is considered the most environment- friendly route of production of hydrogen, which is the most promising alternative to fossil fuels because it is clean, efficient, and renewable [2, 3].

Biological method mainly includes photosynthetic hydrogen production and fermentative hydrogen production. The efficiency of photosynthetic hydrogen production is low and it cannot be operated in the absence of light, while fermentative hydrogen production can produce hydrogen continuously without light. Moreover, compared with photosynthetic hydrogen production, fermentative hydrogen production has higher hydrogen production efficiency, higher hydrogen production stability, higher feasibility for industrialization, simpler control requirement and lower operating costs. Therefore, Dark fermentative hydrogen production is more feasible, widely used and it has been received increasing attention in recent years [4].

*Escherichia coli* is a facultative anaerobe organism, it is characteristic of mixed acid fermentation, it is the best-

characterized bacterium [5]. *E coli* produce hydrogen via its formate hydrogen lyase (FHL) system from formate [6].

This Study has been carried out concerning the effects of temperature and pH on fermentative hydrogen production and to optimize the fermentation factors for improved hydrogen production rate using *Escherichia coli* ATCC 8739 strain and glucose as substrate.

## II. MATERIALS AND METHODS

*E. coli* ATCC 8739 strain was cultured in modified M9 medium containing 2 mL of Pfennig and Lippert's trace element solution [7]. The culture bioreactors were sparged with argon (Ar) gas (99.9%) to ensure that they were completely devoid of O<sub>2</sub> [8]. Glucose was used as carbon source. Batch cultivations were carried out in bioreactors under varying temperatures (25– 40°C) and pHs (5.5- 7.0) at 150 rpm.

The produced gas is measured by water displacement method. It was collected in a graduate glass tube of 150 ml, containing 20% NaOH solution for selective absorption of carbon dioxide, sealed in its top by a septum for gas analysis. The extremity of the tube was directed through a bubbler. The gas composition was confirmed by gas chromatography (SHIMADZU GC-14B) equipped with a thermal conductivity detector (TCD). The injector, the column and the detector temperatures were 100°C, 200°C and 100°C respectively. Argon was used as the carrier gas at a flow rate of 1 l/h [8].

## III. RESULTS AND DISCUSSION

### A. Effect of initial pH

The effect of pH on hydrogen production rate was studied by varying the initial pH of the M9 medium between 5.5 and 7.5. It can be seen from Fig. 1 that the hydrogen production rate varied considerably with pH, it increased with increasing the pH up to pH 6.5 and then the hydrogen production rate was reduced. Experimental data revealed also that the maximum hydrogen production rate was obtained at pH 6.5.

The lower values of hydrogen production rate, were obtained at pHs extremes, higher than 7 and lower than 6, it is may be due to the effect of this pH range on the transport across the cell membrane, on the activity of some cell

biocatalysts, and on the redirection of their metabolic pathways which are competing for pyruvate, in consequence the pH is able to change the composition of fermentation end products that comprise a mixture of fatty volatiles acids, alcohols and hydrogen. Bowles et al [9] and Chittibabu et al [10] reported that, at lower pH, the increased formation of acidic or alcoholic metabolites, which destroys the cell's ability to maintain internal pH [9], it might have resulted in poor hydrogen production and lowering of intracellular level of ATP, thereby inhibiting glucose uptake [10]. So, the pH gradient across the cytoplasmic membrane determines the glucose transport rate. Additionally, Rossmann et al [11] reported that, at alkaline pH the formate excrete out cells through a symport mechanism. Therefore, the intracellular formate concentration reduced and we can conclude that the FHL enzyme complex involved in formate conversion into H<sub>2</sub> and CO<sub>2</sub> could become less active and thus the conversion rate decreased.

The optimum pH of 6.5 observed in this study to maximize hydrogen production rate is in good agreement with the reported values of similar studies using glucose and a pure culture of E coli strain DJT135 [12]. However, Chittibabu et al. [11] reported that pH 6.0 was most suitable for the hydrogen production by E coli strain BL-21. This disagreement in performance of bioprocess with respect to the initial medium pH for enhancing the rate of hydrogen production is dependent on the mechanism of fermentative hydrogen production, on the type of microorganisms and on the experimental conditions.

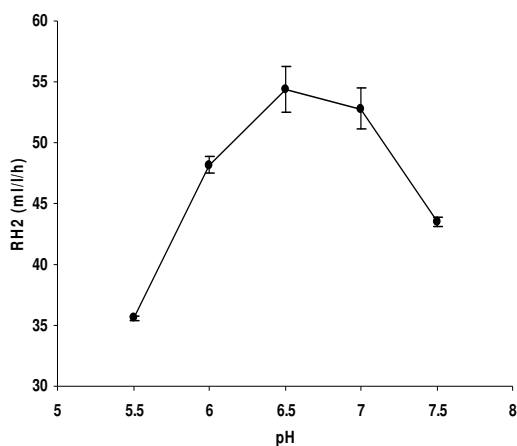


Fig. 1. Effect of pH on hydrogen production rate

### B. Effect of Temperature

In order to determine the optimal temperature for elevating hydrogen production rate by E coli ATCC 8739 from glucose, the effect of temperature ranging from 25°C to 40°C on fermentative hydrogen production was investigated in batch tests as presented in Fig. 2.

The results showed that the hydrogen production rate increased with increasing temperature from 25 °C to 35°C, however, it decreased with further increasing temperature from 35°C to 40°C. The possible reason for the development in bioprocess efficiency with increasing temperature from 25°C to 35°C was that the ability of hydrogen-producing bacteria to degrade substrate and produce hydrogen increased

with increasing temperatures. Moreover, literature shows that lower hydrogen production rate was observed at the temperatures higher than 45°C [13] because some essential enzymes and proteins associated with cell growth or hydrogen production, such as hydrogenase, may be inactivated by an increase in denaturation rate of the enzymes when the temperature gets too high [14, 15].

Maximum hydrogen production rate was obtained with temperature of 35°C, using E coli ATCC 8739, which was also shown using Escherichia coli DJT135 [12].

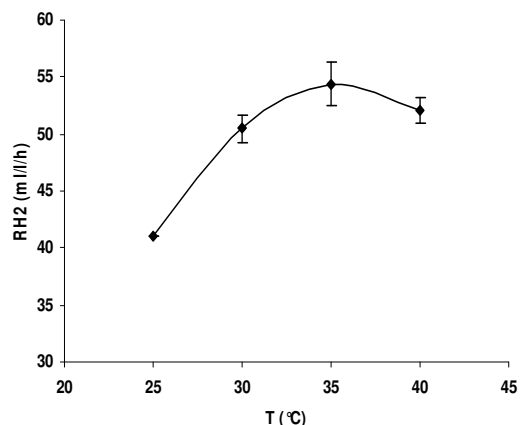


Fig. 2. Effect of temperature on hydrogen production rate

### III.3. COMPARATIVE HYDROGEN PRODUCTION RATES OF DIFFERENT STRAINS OF E COLI

In the present study, the hydrogen production rate of E. coli ATCC 8739 strain was compared with those of various strains of E.coli in batch process (Table I).

TABLE I. HYDROGEN PRODUCTION RATE OF VARIOUS E COLI STRAINS

Strain	Culture and Substrate	Hydrogen Production Rate	References
<i>E coli ATCC 8739</i>	Batch Glucose	56.84 ml/l/h	This work
<i>E coli HD 701</i>	Batch Glucose	52 ml/l/h	[16]
<i>E coli W3110 (K-12)</i>	Batch Glucose and formate	0.3 ± 0.1 mmol/h/g dry cell	[17]
<i>E coli K-12</i>	Batch Glucose and formate	45.8 ± 4.1 mmol/h/g dry cell	[18]

### IV. CONCLUSIONS

This work shows that pH and temperature had a significant influence on the hydrogen production rate. The most suitable pH and temperature for optimal performance of the hydrogen production by E coli ATCC 8739 strain were pH 6.5 and 35°C, respectively. Different E coli strains were compared for their capability in hydrogen production under anaerobic conditions. The experimental data revealed that the

E coli ATCC 8739 strain is a promising hydrogen producer bacterium.

#### REFERENCES

- [1] C. J. Winter, "Into the Hydrogen Energy economy- milestones", *International Hydrogen Energy*, vol. 30, pp. 681 – 685, 2005.
- [2] N. Basak and D. Das, "Photofermentative hydrogen production using purple non- sulfur bacteria *Rhodobacter sphaeroides* OU.001 in annular photobioreactor: A case study" *Biomasse and Bioenergy*, vol. 33, pp. 911-919, 2009.
- [3] R. Y. Li and H. H.P. Fang, "Hydrogen production characteristics of photoheterotrophic *Rubrivivax gelatinosus* L31", *International Journal of Hydrogen Energy*, vol. 33, pp. 974– 980, 2008.
- [4] J. Wang, W. Wan, "Experimental design methods for fermentative hydrogen production: A review", *International Hydrogen Energy*, vol. 34, pp. 235-244, 2009.
- [5] T. Maeda, V. Sanchez-Torres, T. K. Wood, "Enhanced hydrogen production from glucose by metabolically engineered *Escherichia coli*," *Appl Microbiol Biotechnol*, vol. 77, pp. 879–890, 2007.
- [6] T. Maeda, T. K. Wood, "Formate detection by potassium permanganate for enhanced hydrogen production in *Escherichia coli*, *International Journal of Hydrogen Energy*," vol. 33, pp. 2409 – 2412, 2008.
- [7] S. Kim, E. Seola, You-Kwan Ohb, G.Y. Wangc, S. Park, , "Hydrogen production and metabolic flux analysis of metabolically engineered *Escherichia coli* strains," *International Journal of Hydrogen Energy*, vol. 34, pp. 7417- 7427, 2009.
- [8] D. Akroum-Amrouche, N. Abdi, H. Lounici, N. Mameri, "Effect of physico-chemical parameters on biohydrogen production and growth characteristics by batch culture of *Rhodobacter sphaeroides* CIP 60.6," *Applied Energy*, vol. 88, pp. 2130-2135, 2011.
- [9] L.K. Bowles, W.L. Ellefson, " Effects of butanol on *Clostridium acetobutylicum*," *Appl Environ Microbiol*, vol. 50, pp. 1165–70, 1985.
- [10] G. Chittibabu, K. Nath, D. Das, "Feasibility studies on the fermentative hydrogen production by recombinant *Escherichia coli* BL-21," *Process Biochemistry*, vol. 41, pp. 682-688, 2006.
- [11] R. Rossmann, G. Sawers, A. Bock. "Mechanism of regulation of the formate-hydrogenlyase pathway by oxygen, nitrate, and pH: definition of the formate regulon," *Mol Microbiol*, vol. 5, pp. 2807– 14, 1991.
- [12] D. Ghosh, P. C. Hallenbeck, " Fermentative hydrogen yields from different sugars by batch cultures of metabolically engineered *Escherichia coli* DJT135," *International journal of hydrogen energy*, vol. 34, pp. 7979- 7982, 2009.
- [13] J. Wang, W. Wan, " Effect of temperature on fermentative hydrogen production by mixed cultures," *International of Hydrogen Energy*, vol. 33, pp. 5392 – 5397, 2008.
- [14] K-S. Lee, P-J. Lin, J-S. Chang, "Temperature effects on biohydrogen production in a granular sludge bed induced by activated carbon carriers," *International Journal of Hydrogen Energy*, vol. 31, pp. 465 – 472, 2006.
- [15] B. Fabiano, P. Perego, " Thermodynamic study and optimization of hydrogen production by *Enterobacter aerogenes*," *International Journal of Hydrogen Energy*, vol. 27, pp.149–56, 2002.
- [16] M. D. Redwood, L. E. Macaskie, " Atwo-stage, two-organism process for biohydrogen from glucose", *International Journal of Hydrogen Energy*, vol. 31, pp. 1514 – 1521, 2006.
- [17] A. Yoshida , T. Nishimura, H. Kawaguchi, M. Inui , H. Yukawa, "Efficient induction of formate hydrogen lyase of aerobically grown *Escherichia coli* in a three-step biohydrogen production process," *Appl Microbiol Biotechnol*, vol. 74, pp.754–760, 2007.
- [18] E. Seol, S. Kim, S. M. Raj, S. Park, "Comparison of hydrogen-production capability of four different *Enterobacteriaceae* strains under growing and non-growing conditions", *International Journal of Hydrogen Energy*, vol. 33, pp. 5169 – 5175, 2008.